

Bioscene

Bioscene

Volume- 22 Number- 03 ISSN: 1539-2422 (P) 2055-1583 (O) www.explorebioscene.com

Diversity and Cellulase Activity of Chaetomium globosum (Kunze), A Destructive Cellulolytic Fungus

¹Supriya Pandey, ²Shailendra Yadavand, ³Neeraj Srivastava

³Professor and Head ^{1,2,3}Applied Mycology Lab, Department of Botany St. Andrew's (Post-Graduate) College Gorakhpur, Uttar Pradesh, India

Corresponding Author: Neeraj Srivastava

Abstract: Fungi invading objects containing cellulose are called cellulolytic fungi. These fungi produce "cellulolytic enzymes or cellulases", which decompose cellulose (a polysaccharide) into first oligosaccharide and then monosaccharides, and ultimately to glucose molecules. Chaetomium globosum (Kunze) is reported to be a destructive cellulolytic fungus causing damage to our cultural heritage of library books, monuments, paintings, archival materials etc. In the present investigation, diversity of C. globosum in different library books of District - Gorakhpur (located in North- Eastern Uttar Pradesh, India) has been studied. Cellulase activity of C. globosumisolated and cultured has been assayed using Carboxyl Methyl Cellulose (CMC) Agar medium technique. Gram's iodine solution is used as indicator. The Enzyme Index of C.globosum has been reported as 3.6, which shows that it is industrially important fungus, which may be exploited for the production of cellulase enzyme at industrial level. Therefore, this research work strongly supports the concept that screening of cellulase producing ability of C. globosum and other microorganisms is very important.

Keywords: Cellulolytic Fungi, Biodeterioration, Cellulase Activity, Chaetomium globosum, Mesophilic, Enzyme Index (EI).

Introduction

Chaetomium globosum (Kunze) is a mesophilic cellulolytic fungus. It has a vast range of substratum for its growth including plants, soils, debris of plants and animals, papers, textiles and clothes etc. (Sharma et al. 2025). In addition to its saprophytic nature, strain of Chaetomium globosum has been reported to be "Endophytic", assisting in cellulose decomposition of plant cells (Chapman et al. 1975). Its habitat ranges from forest plants to soils of various biomes (Domsch et al. 1980; Liu et al. 2007); to indoor environment including books and wood (Provost et al. 2013). Rarely

some strains of *Chaetomium globosum* are reported to be human allergens, causing Mycosis and neurological infections (Kim *et al.* 2008; Provost *et al.* 2013).

In humid tropical and sub-tropical countries of the world, biodeterioration of various cultural commodities in textiles, papers, wood and leather is a very common phenomenon (Tiano, 2009; Ortega - Morales *et al.* 2019).

Our cultural heritages are heavily damaged and destroyed by these microorganisms (Koestler and Vedral, 1991). The two most important microbes responsible for this biodeterioration are fungi and bacteria. Fungi of various groups play most active role and cause maximum damage to these important objects (Arroyo, 2007).

District Gorakhpur is situated in North - Eastern Uttar Pradesh, India, in foot hills of Himalayas at the bank of Rapti river (coordinates; 26.7637152° N; 83.4039116° E), with average annual temperature of 26°C (79°F); elevation- 75m (246ft). Its high relative humidity and moderate temperature from July to March are very suitable for growth of Mesophilic microbes including fungi (Srivastava, 2007).

Fungi having ability to hydrolyse cellulose (a polymer of glucose) into its simpler units are called cellulolytic fungi (Arnthong et al. 2020). Twenty-six species of these cellulolytic fungi belonging to 15 genera have been reported causing biodeterioration of Webster's Dictionary in Gorakhpur (Srivastava et al. 2011). These fungi by producing cellulase enzymes, cause biodegradation of our cultural heritage containing cellulose, including important library books etc. (Mesquita et al. 2009; Sterflinger et al. 2013). Their activities are supported by high moisture content, moderate light intensity and temperature (Garg et al. 1995; Sterflingeret al. 2012; Coronado-Ruiz et al. 2018).

Biodeterioration of papers and archival material caused by fungi is very common in India (Agrawal, 1995). Lignocellulosic biomass is heavily decomposed by these cellulolytic fungi (Anwar et al. 2014). These fungi cause biodegradation of cellulose containing objects by producing three types of cellulases - "Endo 1,4- β -D-glucanase (EG), Cellobiohydrolase (CBH) or exo 1,4, β -D-glucanse and β -glucosidase" (Teeri, 1997; Bayer et al.1998; Zhang and Lyund 2004; AL-Kharousi et al. 2015). Degradation of cellulose is achieved by the combined effects and actions of these three cellulase enzymes (Qin et al. 2010).

Chaetomium globossum with other cellulolytic fungi has been reported to degrade cellulose, causing damage of important documents. (Pinzari et al. 2006; Michaelsen et al. 2009).

The cellulase producing and decomposing ability of various cellulolytic fungi including *Chaetomium* is reported by several workers (Ames, 1961; Aranyanak, 1995; Al-Kharousi *et al.* 2015).

Exploitation of these cellulolytic fungi and their enzymes at industrial scale is the need of time (Arif et al. 2024). Control of these cellulytic fungi and protection of our cultural heritage in papers and cloths/textiles is a challenging job (Sequeira et al. 2017). However, isolation of these cellulolytic fungi from deteriorated objects and their identification is the initial step. Pure cultures of these fungi are then used to evaluate their cellulolytic activities by calculating their enzyme index (EI).

In the present research work, isolation of cellulolytic fungi from deteriorated samples of papers and cloths/textiles has been done to study the presence of *C. globosum*. Cellulase activity of isolated *C. globosum* has been determined by calculating its enzyme index (EI).

Materials and Methods

1. Isolation of Test Fungus - Chaetomium globosum Kunze

Various sites of District Gorakhpur (such as Government District Library; Central Library of D.D.U. Gorakhpur University and Central library of St. Andrew's Post-Graduate College; Buddha Museum and whole sale market of clothes/textiles) were visited and infested samples of papers and cloths/textiles were collected from these sites.

For isolation, 3 techniques were used – Direct Observation, Standard Blotter Method and Czapek Dox Agar Method.

Six species of *Chaetomium* were isolated and reported from deteriorated samples of papers and cloths/textiles, which were given Isolate/Culture Numbers (**Table - 1**).

Table – 1
Frequencies of Species of Chaetomium Kunze Isolated from
Deteriorated Samples of Papers and Cloths/Textiles

S.	Species of Chaetomium	Isolate/Culture	DO	SBM	CDA
No.	Isolated	No.			
1.	Chaetomium	SP/AML/0122	+	+	++
	dolichotrichum Ames				
2.	C. funiculosum Corda	SP/AML/0123	+	+	
3.	C. globosum Kunze	SP/AML/0124	+++	++	+++
4.	C. strumariumMinter	SP/AML/0125	+	+	++
5.	Chaetomium sp. (US) - 1	SP/AML/0126	+	-	+
6.	Chaetomium sp. (US) - 2	SP/AML/0127	+	-	-

+++ : Most Frequent DO : Direct Observation

++ : More Frequent SBM : Standard Blotter Method

+ : Less Frequent CDA : Czapek Dox Agar
- : Absent US : Unidentified Species

As it is evident from **Table – 1**, maximum frequency observed was of *C. globosum*, which was selected as the test fungus. It was cultured on Potato Dextrose Agar (PDA) Medium.

The pure culture of isolated *C.globosum* (Isolate/Culture No. SP/AML/0124) was deposited in NFCCI (National Fungal Culture Collection of India), Pune, Maharashtra, India to get Accession No. (Accession. No. – NFCCI5963). Also, the pure culture of *C. globosum* was procured from ITCC (Indian Type Culture Collection), Indian Agricultural Research Institute (IARI), New Delhi (ITCC. No. - 3680).



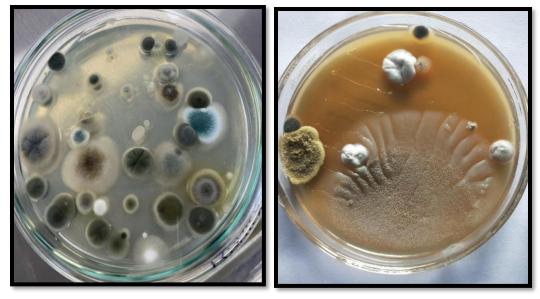


Shakya Buddha, Thanka (1875 AD, TIBBET) infested by Cellulolytic fungi

Infested page of "Hindu Dharmkosh"

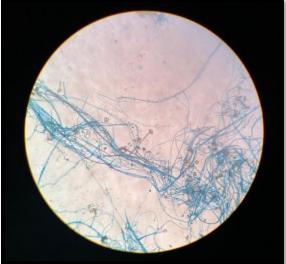


Damaged Book entitled "SUKH SAGAR" Isolation of Cellulolytic Fungi (1856)



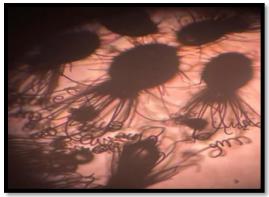
Mixed Culture of Cellulolytic Fungi Isolated





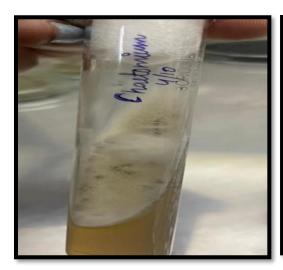
Pure Culture of Chaetomium globosum

Mycelium of Chaetomium globosum





Spores of Chaetomium globosum





Slant of Chaetomium globosum for Pure Culture of Chaetomium globosum

Deposition in NFCCI, Pune procuredfrom ITCC, IARI, New Delhi

2. Cellulase Assay of Chaetomium globosum

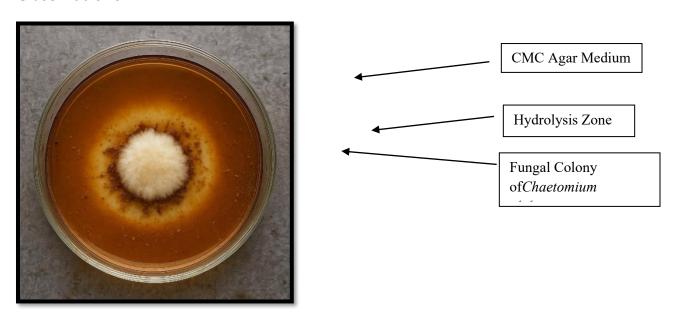
Cellulase assay of the test fungus C.globosum was done by using 1% Carboxyl Methyl Cellulase Agar medium (CMC AM).

Carboxyl Methyl Cellulose serves as source of carbon and Gram's iodine solution as indicator to observe cellulose hydrolysis zone (Coronado - Ruiz et al. 2018). In this process of qualitative determination of cellulase activity, the Gram's iodine, react with cellulose as well as its decomposed components. The unhydrolyzed and intact cellulose holds the colour of Gram's iodine solution. However, that cellulose which is hydrolysed by fungal cellulase enzymes reflects 'clear zone' (seen as pale-yellow zone). The enzymatic index of *C.globosum* was calculated as-

EI of
$$C. globosum = \frac{Diameter of Hydrolysis Zone}{Diameter of Colony Growth of $C. globosum$$$

To evaluate E.I., 6mm. diameter disc of pure culture of C. globosum was cultured on 1% CMC Agar Medium and was incubated for 7 days at 28 ± 2 °C temperature. The antibiotic used was Streptomycin. Gram's Iodine solution was used as indicator. Afterwards, this culture was washed with water. Colony diameter of C. globosum and clear zone around the fungal disc were measured to calculate enzyme index (Coronado-Ruiz; 2018).

Observations



Cellulase Assay of Chaetomium globosum

- 1. Diameter of hydrolysis zone (Diameter of colony + clear zone) = 36mm.
- **2.** Diameter of colony growth of C. globosum = 10mm

Therefore:

EI of *C. globosum* =
$$\frac{36 \text{ mm.}}{10 \text{ mm.}}$$
 = 3.60

Result and Discussion

Six species of Chaetomiumhave been isolated and cultured from deteriorated samples of papers and clothes/ textiles. Of these 6 species, 4have been identified as Chaetomium dolichotrichum Ames, C. funiculosum Corda, C. globosum Kunze and C. strumarium Minter. However, two unidentified species (US) were also isolated and were given culture numbers for future studies. Of all these 6 species, C. globosum was reported to be the most frequent species collected from all the samples.

Enzymatic index (EI) of C. globosum has been calculated as 3.60

The results of present work clearly shows that *C.globosum* is efficient in cellulase enzyme production and cellulase producing ability of this species can be exploited at industrial level. Also, the objects containing cellulose, such as books, archives, cloth and textiles *etc.* could be a good source to explore and isolate cellulase producing fungi like *C.globosum etc.*

Acknowledgements

We are thankful to NFCCI (National Fungal Culture Collection of India), Agharkar Research Institute, Pune (Maharashtra) for deposition of pure culture of *Chaetomium globosum* to provide Accession Number and to ITCC (Indian Type Culture Collection), Indian Agricultural Research Institute (IARI), New Delhi for supply of pure culture of *Chaetomium globosum*. We are also thankful to the Principal, St. Andrew's College, Gorakhpur for providing infrastructural lab. facilities. Prof. Neeraj Srivastava is thankful to Uttar Pradesh State Higher Education Council, Lucknow, U.P., India for financial support under "Research and Development Project for University and College Teachers".

Authors Contributions

All authors contributed significantly to the final manuscript.

Statements and Declarations: Authors declares no conflict of Interest.

References:

- 1. AL-Kharousi, Moza Mohammed; Sivakumar, Nallusamy and Elshafie Abdulkadir (2015). Characterization of cellulase enzyme produced by Chaetomium sp. isolated from books and archives. Eurasia J Biosci. 9, 52-60.
- 2. Ames, L.M. (1961). A Monograph of the Chaetomiaceae. No. 2. The United Army Research and Development Series, (1961).
- 3. Anwar, Z; Gulfaz, M; Irshad, M. (2014). Agro-industrial lignocellulosic biomass a key to unlock the future bio-energy: A brief review. J. Radiat. Res. Appl. Sci.7: 163–173.
- 4. Aranyanak, C. Microscopical Study of Fungal Growth on Paper and Textile. (1995). In:Biodeterioration of Cultural Property 3. C. Aranyanak and A. Singhasiri. Eds. Proceedings of the 3rd International Conference of Biodeterioration of Cultural Property, Bangkok, (1995):83-102.
- 5. Arif, S; M'Barek H.N.; Bekaert, B; Aziz, M.B.; Diouri, M; Haesaert, G.; Hajjaj H. Lignocellulolytic Enzymes Production by Four Wild Filamentous Fungi for Olive Stones Valorization: Comparing Three Fermentation Regimens. J Microbiol Biotechnol. (2024) May 28;34(5):1017-1028.
- Arnthong, J.; Siamphan, C.; Chuaseeharonnachai. C.; Boonyuen, N; Suwannarangsee, S. (2020). Towards a Miniaturized Culture Screening for Cellulolytic Fungi and Their Agricultural Lignocellulosic Degradation. J Microbiol Biotechnol. 30 (11):1670-1679.
- 7. Arroyo, Irene. (2007). The role of fungi in the deterioration of movable andimmovable cultural heritage. E-Conservation Magazine, Spain:40-50.
- 8. Bayer, E.A.; Chanzy H.; Lamed, R.; Shoham, Y. (1998) Cellulose, cellulases and cellulosomes. Current Opinion in Structural Biology 8: 548-557.
- 9. Chapman, E.; Fergus, C. (1975). "Germination of Ascospores of Chaetomium lobosum". Mycologia **67**: 1048-1052.
- 10. Coronado-Ruiz, C.; Avendaño, R.; Escudero-Leyva, E.; Conejo-Barboza, G.; Chaverri, P.; Chavarría, M. (2018). Two new cellulolytic fungal species isolated from a 19th century art collection. Sci. Rep. 8: 7492: 1-9.
- 11. Domsch, K.H.; Gams, W.; Anderson, T. (1980). "Compendium of Soil Fungi" London, New York: Academic Press.
- 12. Garg, K.L.; Kamal, K.; Mishra, A.K. (1995). Role of fungi in the deterioration of wall paintings. Sci. Total Environ. 167: 255–271.
- 13. Kim, D.; Lee, M.; Suh, M.; Ha, G.; Kim, H.; Choi, J. (2008). "Onychomycosis Caused by Chaetomium globosum". Ann Dermatol **25**(2): 232-236.
- 14. Koestler, R.J.; Vedral, J. (1991). Biodeterioration of Cultural Property: A Bibliography. International Biodeterioration. 28: 229-30.
- 15. Liu, Z.; Yang, Q.; Ma, J. (2007). "A heat shock protein gene (hsp22.4)

- from Chaetomium globosum confers heat and Na2CO3 tolerance to yeast". Appl Microbiol Biotechnol **77**: 901-908.
- 16. Mesquita, N.; Portugal, A.; Videira, S.; Rodri'guez-Echeverri' S.; Bandeira, A.M.L.; Santos, M.J.A.; Freitas, H. (2009). Fungal diversity in ancient documents. A case study on the archive of the University of Coimbra. Int. Biodeter. Biodegr. 63: 626–629.
- 17. Michaelsen, A.; Piñar, G.; Montanari, M.; Pinzari, F. (2009). Biodeterioration and restoration of a 16th-century book using a combination of conventional and molecular techniques: A case study. Int. Biodeter. Biodegr. 63: 161–168.
- 18. Ortega-Morales, Otto; Jorge, Luis Montero-Muñoz; José, Antonio Baptista Neto; Iwona, B. Beech; Jan, Sunner; Christine, Gaylarde. (2019). Deterioration and microbial colonization of cultural heritage stone buildings in polluted and unpolluted tropical and subtropical climates: A meta-analysis. Int. Biodeter. Biodegr. 143, 104734.
- 19. Pinzari, F.; Pasquariello, G.; De Mico, A. (2006). Biodeterioration of paper: a SEM study of fungal spoilage reproduced under controlled conditions. Macromol. Symp. 238: 57–66.
- 20. Provost, N.; Shi, C.; She, Y.; Cyr, T.; Miller, D. (2013). "Characterization of an antigenic chitosanase from the cellulolytic fungus Chaetomium globosum". Medical Mycology **51**:290-299.
- 21. Qin, Y; He, H.; Li, N.; Ling, M; Liang, Z. (2010) Isolation and characterization of a thermostable cellulase-producing Fusarium chlamydosporum. World Journal of Microbiology and Biotechnology 26: 1991-1997.
- 22. Sequeira, S.; Phillips, A.; Cabrita, E.; Macedo, M. (2017). Antifungal treatment of paper with calcium propionate and parabens: Short-term and long-term effects. Int. Biodeterior. Biodegradation. 120: 203–215.
- 23. Sharma, S.; Pandey, S.; Kulshreshtha, S.; Dubey, M. (2025). Biology and Application of Chaetomium globosum as a Biocontrol Agent: Current Status and Future Prospects. Microorganisms: 13:01-19.
- 24. Srivastava, Mamta; Arya, Mithilesh Kumar and Srivastava, Neeraj. (2011). Cellulolyticfungi causing biodeterioration of Webster's dictionary in Gorakhpur. International Journal of Biological Technology Vol. 2 (Special Issue). 216-220.
- 25. Sterflinger, K; Pinzari, F. (2012). The revenge of time: fungal deterioration of cultural heritage with particular reference to books, paper and parchment. Environ. Microbiol. 14: 559–566.
- 26. Sterflinger, K.; Piñar, G. (2013). Microbial deterioration of cultural heritage and works of art tilting at windmills. Appl. Microbiol. Biotechnol. 97: 9637–9646.
- 27. Teeri, Tuula T. (1997). Crystalline cellulose degradation: new insight into the

- function of cellobiohydrolases. Trends in Biotechnology. 15 (5): 160-167.
- 28. Tiano, Piero. (2009). Biodegradation of Cultural Heritage: Decay Mechanisms and Control Methods. ResearchGate Article. Content uploaded by Piero Tiano. www.researchgate.net.
- 29. Zhang, YHP; Lynd, L.R. (2004). Toward an aggregated understanding of enzymatic hydrolysis of cellulose: Noncomplexed cellulase systems. Biotechnology and Bioengineering. 88: 797-824.