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Morpho-Taxonomy and Phytopathology of *Cercospora* Species Invading Medicinal Plants in North-Eastern Uttar Pradesh, India

¹Dilip Kumar Tripathi; ²Fakhare Do Alam & ³Neeraj Srivastava

³Professor and Head

^{1,2,3} Applied Mycology Lab, Department of Botany, St. Andrew's (Post-Graduate) College, Gorakhpur, Uttar Pradesh, India

Corresponding Author: **Neeraj Srivastava**

Abstract: North-Eastern Uttar Pradesh, India has rich biodiversity of plants. *Cercospora* Fres. is a phytopathogenic fungus causing diseases in different plants of economic importance. In the present investigation, during survey of villages and forests of Gorakhpur district, three phytopathogenic species of *Cercospora* have been collected and reported - *Cercospora achyranthina*, *C. cocciniae* and *C. papaya*, infecting three medicinally important plants - *Achyranthes aspera*, *Coccinia grandis* and *Carica papaya*. Morpho-taxonomy of these three species of *Cercospora* including camera lucida drawings have been done. Phytopathological investigations have also been performed including reductions in photosynthetic areas of leaves. It is concluded that these *Cercospora* species affect the productivity as well as medicinal properties of these medicinally important plants.

Keywords: Follicolous Fungi, Morpho-taxonomy, Phytopathology, Medicinal Plants, *Cercospora*

Introduction

District Gorakhpur of North-Eastern Uttar Pradesh, India is located at the Rapti riverside, in the foot hills of Himalayas (Geographical location - 26.7637152° N; 83.4039116° E; average annual temperature of 26 - 27°C; elevation- 246ft). Its climatic conditions (high relative humidity and moderate temperature) from July to November are very suitable for plant pathogenic fungi (Srivastava et al., 2011).

District Gorakhpur has a rich plant biodiversity including forest flora as well as plants of agricultural, horticultural and medicinal importance (Srivastava, 2007). Its humid and subtropical climate supports growth of various pathogens including fungal and bacterial pathogens etc. (Yadav et al., 2025).

The fungus genus *Cercospora* Frasenius ex Fuckel has been reported to be a phytopathogenic fungus (Systematic position: Kingdom- Fungi; Phylum- Ascomycota; Class- Deuteromycetes; Order- Moniliales; Family- Dematiaceae). It causes diseases in Angiosperms (Dicot and Monocot plants), Gymnosperms and Ferns etc. (Pollack, 1987). Its symptoms include leaf spots as well as necrosis of

flower, fruits, seeds and pedicels etc. (Agrios, 2005). Some species of *Cercospora* are used in biological control as these are hyperparasites on plant pathogenic fungi (Shin and Kim, 2001; Morris and Crous, 1994).

The monograph of *Cercospora* (Chupp, 1954) provided a rich information about various species of *Cercospora*. The genus *Cercospora* was further reclassified into numerous species (Deighton, 1967, 1971, 1973, 1974, 1976, 1979, 1983). Presence of conidial scar was taken as an important taxonomic criterion by Deighton as well as other mycologists too (Pons and Sutton, 1988; Braun, 1988 a, b, 1989, 1990, 1993).

Crous and Braun (2003) compiled over 3000 species of *Cercospora*. Thirumalachar and Chupp (1948) described and commented on Indian *Cercosporae*, and a number of plant pathogenic species of cercosporoid fungi including *Cercospora* have been reported from North-Eastern Uttar Pradesh, India by Kamal and his co-workers (Kamal, 2010; Sinha et al., 2022; Srivastava et al., 1994; Srivastava et al., 1994; Srivastava et al., 1995; Srivastava et al., 1995; Srivastava and Morgan Jones, 1996; Bhalla et al., 1996; Srivastava et al., 2007).

Medicinal Plants under Study

Three phytopathogenic species of *Cercospora* have been reported on medicinally important plants - *Achyranthes aspera*, *Carica papaya* and *Coccinia grandis*.

(1). *Achyranthes aspera* L.

It is a weed of Family - *Amaranthaceae* (Local name Chichira/ Chirchita/ Apamarga/ Latjeera). It is distributed throughout India and abundantly found in district Gorakhpur and surrounding areas. Different plant parts are used as traditional medicine in Ayurveda, Unani and Kabiraji treatment systems to cure different diseases like Oedema/ Edema (Nadkarni, 1954), Arthritis, Leprosy, Heart and Kidney diseases and Asthma (Singh, 1995), Piles, Scorpion and Snakebites, Diabetes, Skin diseases (Jayaveera, 1982; Nadkarni, 1954), Malaria, Cough and fever; Fistula and nasal infections, dysentery, cold and tooth ache (Borthakur and Gowswami, 1995; Aziz et al. 2005); Gynaecological problems (Khan and Khan 2006; Shukla et al. 2008) and Stomach pain (Ghani et al. 2003). The phytochemistry of this plant has proved presence of various medicinally important compounds like Alkaloids (Kapoor and Singh 1966), Flavonoids (Sinha and Dogara, 1985), Achyranthin (Basuet al. 1957; Basu et al. 1997; Bhom, 1992; Kapoor and Singh, 1966), Saponins (Batta and Rangaswami, 1973; Shesadri et al. 1981; Srivastava et al. 2002), Glycosides (Arunkumar et al. 2010), Sterols and Ecdysterone (Benerji et al. 1971; Benerji and Chadha 1971; Ikan et al. 1971). These phytochemicals provide pharmacological importance to this plant. This plant has been studied in detail for its pharmacological and phytochemical importance (Krishnaveni and Thakur, 2006; Mali et al. 1990).

Achyranthes aspera plant is medicinally so important that its phyto-pathological investigations have been done by different plant pathologists. Various fungal,

bacterial, phytoplasma, viral and nematode diseases have been reported, (Jairaman, 1988; Raj et al. 2009; Raman et al. 2007).

(2). *Carica papaya* L.

Carica papaya (English- Papaya; Local name - Papeeta) is a tropical fruit tree of Family -Caricaceae. It is well distributed in tropical and subtropical countries of the world including India. According to a report of the year 2020, India is the largest producer of papaya in the world. It is considered as a magic plant because of its nutritional benefits and traditional and modern medicinal uses in disease management etc. Papaya leaves and fruits (unripe and ripe) are rich in phytonutrients (Alara et al. 2020; Karunamoorthiet al. 2014; Saeed et al. 2014). It is used in traditional (folk) medicines (De Oliveira and Vitória, 2011; Starley et al. 1999). The nutraceutical significance of papaya fruits includes presence of antioxidant and immunostimulant compounds, Vitamin - C, Organic acids, Glucose etc. (Begum, 2014). The milky latex of Papaya is used to treat dyspepsia, burns, whooping cough and diarrhea (Reed, 1976). The unripe fruits of papaya have been reported to cure ulcers, menstrual problems and impotency (Elizabeth, 1994). In modern medicine and pharmacology, the leaf extract of papaya is used to treat various human diseases and lethal viruses (Sharma et al., 2022), It contains various types of alkaloids, tannins, glycosides, steroids, saponins and flavonoids etc. (Hu et al. 2012; Patil et al. 2014).

Papaya plant is infected by various plant pathogens including fungi, bacteria, nematodes and viruses etc. (Fatima et al. 2006; Rahman et al. 2008; Srivastava and Singh, 2022; Umer et al. 2022).

(3). *Coccinia grandis* L.

Coccinia grandis (English name - Ivy Gourd; Hindi Name - Kundaroo) is a medicinally important plant of Family-Cucurbitaceae. It is well distributed in tropical and subtropical countries of the world including India (Munir and Qureshi, 2018) and a popular vegetable of district Gorakhpur and surrounding areas.

This plant has been used as traditional herbal medicine to cure various diseases like diabetes (Akhtar et al. 2021; Neamsuvan et al. 2018; Packirisamiet al. 2018; Rahman et al. 2023); as antimicrobial and insecticidal medicine (Mahendrarajah et al. 2020); in hypertension and fever (Uddin et al. 2019); treatment of piles (Kunwar et al. 2012); hyperglycemia (Munir and Qureshi, 2018); in spleen problem and abdominal pain (Bussman et al. 2021); jaundice (Morvin Yabesh et al. 2014) etc. Different parts of this plant like roots, shoots, leaves, flowers and fruits etc. have medicinal properties (Neamsuvan et al. 2018; Waisundara et al. 2015).

Various types of bioactive compounds of *Coccinia grandis* are responsible for its pharmacological activities, such as Alkaloids (Kondhare and Lade, 2017), Senecionin and Cathinone etc. (Chanda et al. 2020), Alkanes like Tritetracontane etc. (Jayalakshmi et al. 2019).

Materials and Methods

Villages and forest areas of district Gorakhpur were visited for collection of diseased plant samples. The infected leaves of *Achyranthes aspera*, *Carica papaya* and *Coccinia grandis* showing disease symptoms were collected, pressed and dried to make permanent Herbarium for depositing in Applied Mycology Lab. as Isotype. Herbarium Cryptogamae Indiae Orientalis (HCIO), IARI, New Delhi was visited to study diseased specimens deposited. The Phytopathology of fungi was studied with special reference to reduction in photosynthetic areas of leaves, which was compared with the leaf areas of healthy leaves.

Morpho-taxonomy, photomicrography and Camera Lucida drawings of *Cercospora* species associated with these three medicinally important plants were done from fresh and dried leaves by using compound microscope.

Observations and Results

Three species of *Cercospora* invading leaves of these three medicinally important plants were described and illustrated as follows:

(1). *Cercospora achyranthina* Thirum and Chupp

on leaves of *Achyranthes aspera* L. (Family-Amaranthaceae).

(i). Leaf Symptoms: Spots are amphigenous (present on adaxial and abaxial, both surfaces), circular to angular or irregular in shape, 2.0 mm to 6.0 mm in diameter (in advanced stage of infection, spots coalesce to form larger spots of 10-12 mm in diameter), light to dark brown in colour with creamish center and reddish-brown margin. Older leaves show dark brown spots with a halo.

(ii). Morpho-taxonomy of Pathogen:

Vegetative Mycelium: Internal, intercellular, branched, septate and each cell is uninucleate.

Stromata: Sub-stomatal formed by aggregation of vegetative hyphae; small in size (10-20 μ m in diameter).

Conidiophore: These are produced in fascicles of 4-6, arising from stromata and emerging out through stomata of leaf; erect to curved and geniculate and sinuous; unbranched, septate (septa 2-10), olivaceous brown in colour, conidiogenous cells are integrated, length is 10 to 40 μ m., clear scars are present on bendings corresponding to hilum of conidia.

Conidia: Solitary, acicular (needle shaped), slightly obclavate-cylindrical, hyaline, thin and smooth walled, apex acute, base truncate, 30 -150 μ m long and 2-5 μ m wide, septate. Each conidium has thick and dark hilum at the base, corresponding to scar on conidiophore.

Holotype: HCIO No. 50223; Isotype: SAC/AML/NS10001

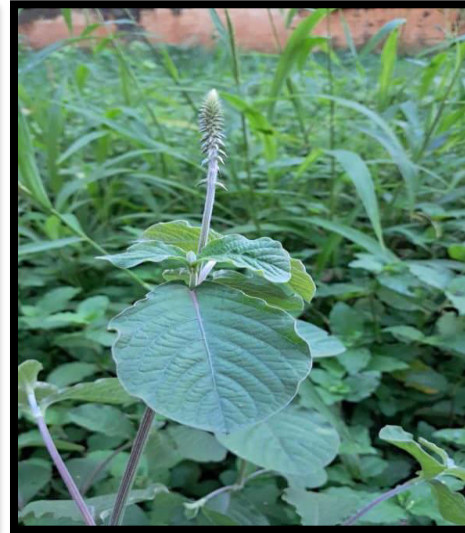
(iii). **Phytopathology of *Cercospora achyranthina*:** The leaf spots on diseased leaves were measured and compared with the leaf areas of healthy leaves. In general, this fungus causes approximately 45-50% reduction in the photosynthetic areas of leaves.

Plate – 1

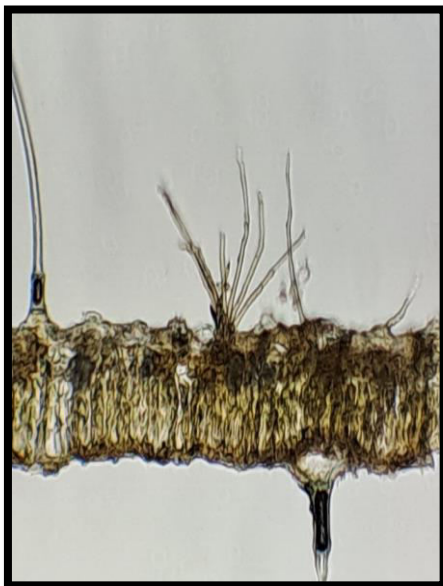
***Cercospora achyranthina* Thirum and Chupp on *Achyranthes aspera* L.**



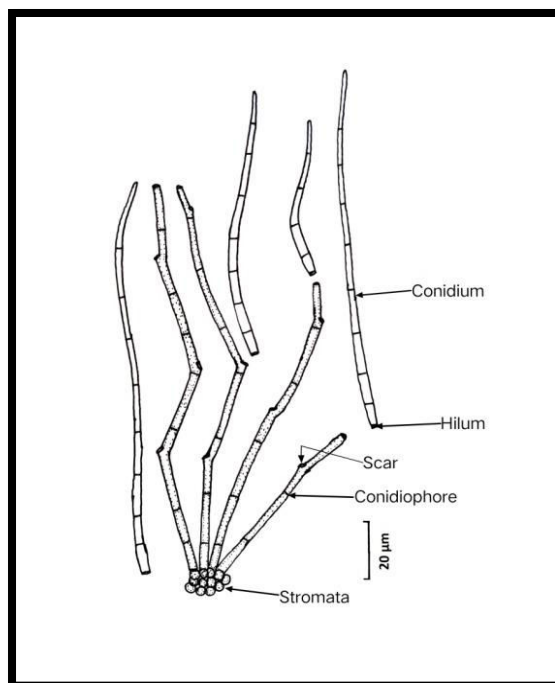
**Diseased Leaf
(Adaxial Surface with Leaf Spots)**



Healthy Leaf



Photomicrographs of V.S. Leaf showing Conidiophores & Conidia



**Camera Lucida Drawings
(Stromata, Conidiophore, Conidia)**

(2). *Cercospora papaya* Hansf

on leaves of *Carica papaya* (Family – Caricaceae).

(i). Leaf Symptoms: Leaf spots amphigenous (present on adaxial and abaxial, both surfaces), 3-6mm in diameter and scattered, circular to sub-circular in shape, creamish to light brown to dark brown in colour, margins dark brown. In advance infection, leaf spots coalesce to form larger spots of 8 to 10 mm in size.

(ii). Morpho-taxonomy of Pathogen:

Vegetative Mycelium: Internal, intercellular branched, septate and uninucleate.

Stromata: These are formed by aggregation of mycelia beneath the epidermis. 15-30μm in diameter, made of sub-globular to irregular brown cells.

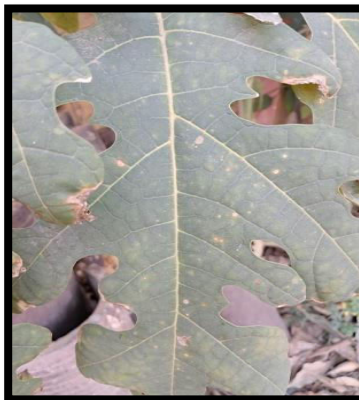
Conidiophores: Produced in divergent fascicles of 5-15 (45-200 μm long and 3-5 μm wide), emerging out through stomata, olivaceous brown in colour, straight to slightly curved and geniculate, scar is present in each bent corresponding to hilum of conidium.

Conidia: Solitary, acicular, obconically truncate at the base, straight to curved, hyaline, septate (15-40 septa), thick and dark hilum (1 to 2 μm in diameter) is present at the base of conidium.

Holotype: HCIO No. 23908; Isotype: SAC/AML/NS 10002

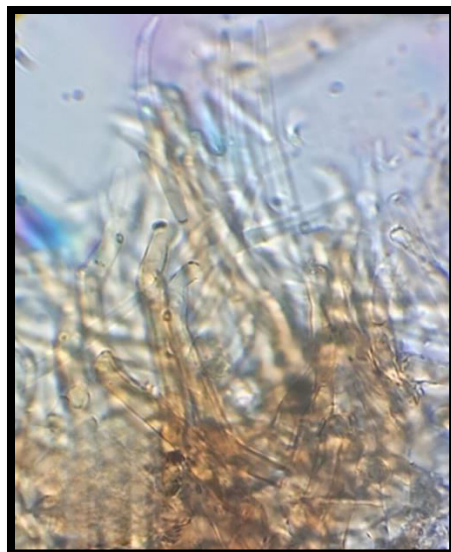
(iii). Phytopathology of *Cercospora papayae*: The areas of leaf spots are measured and compared with total leaf areas of disease-free healthy leaves. It is estimated that this fungus causes approximately 25 to 30% reduction in the photosynthetic areas.

Plate 2:
Cercospora papayae* Hansf on *Carica papaya

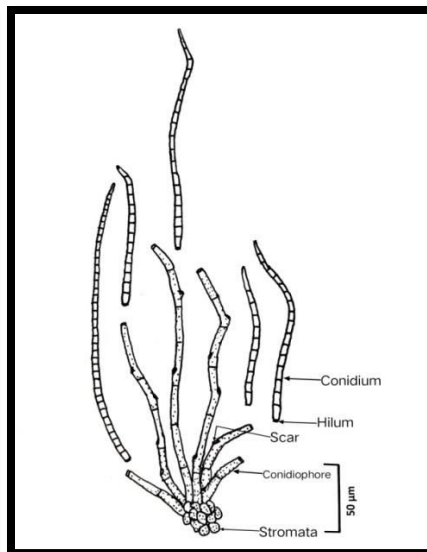


Diseased Leaves
(Adaxial Surface with Leaf Spots)

Healthy Leaf



Photomicrograph of Conidiophores
and Conidia



Camera Lucida Drawings
(Stromata, Conidiophore, Conidia)

(3). *Cercospora cocciniae* Munjal, Lall and Chona

On leaves of *Coccinia grandis* (Family – Cucurbitaceae)

(i). Leaf Symptoms: Leaf spots are 2-6 mm in diameter and amphigenous (present on adaxial and abaxial, both surfaces), circular to sub-circular in shape, light yellow to creamish in colour and whitish in the centre, the margin is light brown to dark brown.

(ii). Morpho-taxonomy of Pathogen:

Vegetative Mycelium: Internal, intercellular, branched, septate and uninucleate.

Stromata: 20-24 μm in diameter, sub-globular in shape and light brown to dark brown in colour.

Conidiophore: Simple, straight, light yellow to brown, $48-82 \times 4-7 \mu\text{m}$ in diameter, 6 to 12 in number, branched and geniculate. Conidiophores form loose fascicles and come out on the leaf surface through stomata. Thick and dark scars are present on each bending, corresponding to the hilum of conidia. Scars are also present, one on each apex of conidiophore. Conidiogenous cells are holoblastic.

Conidia: 70-240 μm long and 1-4 μm wide, produced solitary on each bending of conidiophores, sub acicular to obclavate in shape, straight, colourless, septate (septa 5 to 16). Thick and dark hilum is present at the base of each conidium corresponding to scar on conidiophore; rim of hilum is 1 to 3 μm in diameter.

Holotype: HCIO-50121; Isotype: SAC/AML/NS- 10003.

(iii). Phytopathology of *Cercospora cocciniae*

The spots of diseased leaves were measured and compared with the total leaf areas of healthy leaves. In general, this fungus causes 35-45% reduction in photosynthetic areas of leaves.

Plate 3

***Cercospora cocciniae* Munjal, Lall and Chona on *Coccinia grandis* L.**



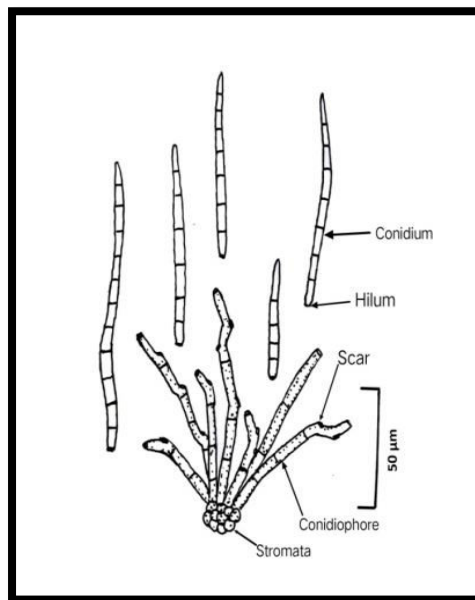
Diseased Leaves
(Adaxial Surface with Leaf Spots)



Healthy Leaf



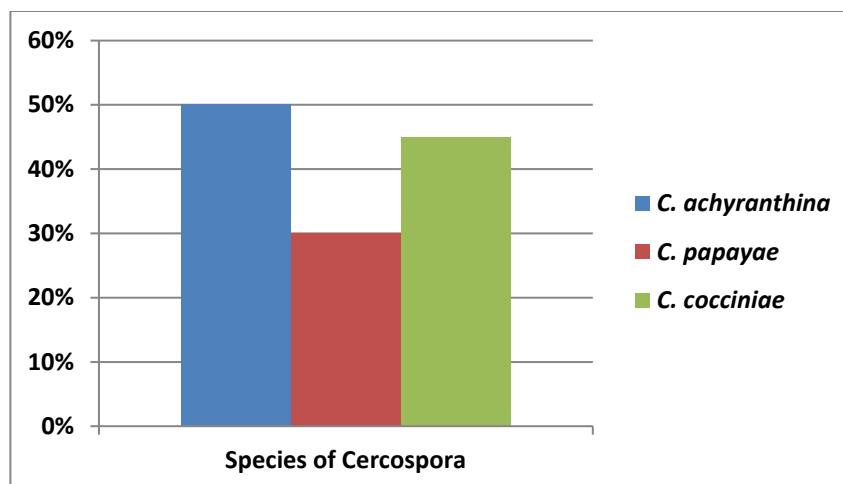
Photomicrographs of Conidiophores and Conidia Single Conidium with Hilum Rim



**Camera Lucida Drawings
(Stromata, Conidiophore, Conidia)**

Discussion

As it is evident from present investigations, species of *Cercospora* produce smaller to larger leaf spots, thereby causing reduction in the photosynthetic areas of leaves and consequent reduction in the productivity of host plants. This results into less production of medicinally/pharmacologically important compounds and secondary metabolites of plants. Therefore, the infected and diseased plants have less medicinal importance as compared to healthy plants.



Per cent Reduction in Photosynthetic Areas of Leaves

Cercospora achyranthina is showing maximum reduction in photosynthetic areas of leaves of *Achyranthes aspera* (45-50%), followed by *C. cocciniae* (35-45%). *Cercospora papayae* shows minimum reduction in photosynthetic area of *Carica papaya* (25-30%).

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References:

1. Agrios, G.N. (2005). Plant Pathology, 5th ed. Academic Press, New York.
2. Akter, K.M.; Sajib, N.H.; Kang, D.M.; Ahn, M.J.; Uddin, S.B. (2021). Ethnomedicinal study of plants in Begumganj, Noakhali, Bangladesh. Nat. Prod. Sci. 27(4):217–227.
3. Alara, O.R.; Abdurahman, N.H.; Alara, J.A.(2020). *Carica papaya*: Comprehensive overview of the nutritional values, phytochemicals and pharmacological activities. Adv. Tradit. Med. 22:1–31.
4. Aminabee, S.; Deepthi, V.; Haribabu, S.; Karthik, S.; Tejaswi, M.; Naga Sai, P.; Lakshmana Rao, A.; Pradesh, A.(2022). Anticancer activity of isolated

- constituents from *Coccinia grandis* by sulphorhodamine (Srb) Assay on Du-145 and Pc-3 Cell Lines. *Eur. J. Mol. Clin. Med.* 09(08):84-102.
5. Arunkumar, S.; Muthuselvam M.; Rajasekaran, R. (2010). Analysis of phytochemical constituents and antimicrobial activity of some Southern India medicinal plants. *Journal of Pharmacy Research.* 3(8):1841- 1843.
 6. Aziz, A.; Rahman, M.; Mondal, A.K.; Muslim, T.; Rahman, A.; Quader, A. (2005). 3- Acetoxy-6 benzoyloxyapangamide from *Achyranthes aspera*. *Dhaka Univ. J. Pharm. Sci.* 4(2):113-116.
 7. Banerji, A.; Chadha, M.S. (1970). Insect moulting hormone from *Achyranthes aspera* Linn. *Phytochemistry.* 9:1671.
 8. Banerji, A.; Chintalwar, G.J.; Joshi, N.K.; Chadha, M.S. (1971). Isolation of ecdysterone from Indian plants. *Phytochemistry.* 10(9):2225- 2226.
 9. Basu, N.K. (1957). The chemical constitution of Achyranthine. *J. Proc. Inst. Chem.* 29: 73-76.
 10. Basu, N.K.; Neogi, N.C.; Srivastava, V.P. (1957). Biological investigation of *Achyranthes aspera* Linn. and its constituent achyranthine. *J. Proc. Inst. Chem.* 29:161-165.
 11. Basu, N.K.; Singh, H.K.; Aggarwal, O.P. (1957). A chemical investigation of *Achyranthes aspera* Linn. *J. Proc. Inst. Chem.* 29:55-58.
 12. Bhalla, K.; Srivastava, N.; and Kamal. (1996). New species of *Mycovellosiella* from North-Eastern Uttar Pradesh, India. *Mycol. Res.* 100(11):1333-1336.
 13. Bhatti, D.S.; Gupta, D.C.; Dahiya, R.S.; Malhan, I. (1974). Additional hosts of the root-knot nematode, *Meloidogyne javanica*. *Current Science.* 43(19):622-623.
 14. Bhom, K.H. (1992). *Achyranthes*, In: Liersch R, Haensel R, Keller K, Rimpler H, Schneider G (Eds.). *Hagers Handbuch der Pharmazeutischen Praxis* VSpringer-VerlagBerlin. 54-59.
 15. Borthakur, S.K.; Gowswami, N. (1995). Herbal remedies from Dimoria of Kamrup district of Assam in northeastern India. *Fitoterapia.* 66:333-340.
 16. Braun, U.; (1988) b. Studies on *Ramularia* and allied genera (II). *Nova Hedwigia.* 47:335-349.
 17. Braun, U. (1989). *Cercospora*-like fungi on *Cassia*. *International Journal of Mycology and Lichenology.* 4:191-204.
 18. Braun, U. (1990). Studies on *Ramularia* and allied genera III. *Nova Hedwigia.* 50:499-521.
 19. Braun, U. (1993). Taxonomic notes on some species of *Cercospora* complex (III). *Mycotaxon.* 48:275-298.
 20. Braun, U. (1988) a. Studies on *Ramularia* and allied genera (I). *International Journal of Mycology and Lichenology.* 3:271-285.
 21. Bussmann, R.W.; Paniagua-Zambrana, N.Y.; Njoroge, G.N.; (2021). *Coccinia grandis* (L.) Voigt cucurbitaceae In: *Ethnobotany of the Mountain Regions of Africa.* Springer International Publishing. pp. 309-311.

22. Chanda, J.; Mukherjee, P.K.; Kar, A.; Maitra, P.K.; Singha, S.; Halder, P.K.; Gajbhiye, R.; Vishnuvardh, R. (2020). LC-QTOF-MS-based metabolite profiling and evaluation of α -glucosidase inhibitory kinetics of *Coccinia grandis* fruit. *Biomed. Chromatography. BMC.* 34(12):4950.
23. Choudhury, S.N.; Paul, S.B.; De, B. (2013). A bioactive alkaloid from the fruit of *Coccinia grandis*. *Asian J. Chem.* 25(17):9561–9564.
24. Chupp, C. (1954). A monograph of the fungus genus *Cercospora*. Published by the author, Ithaca, New York.
25. Crous, P.W.; Braun, U. (2003). *Mycosphaerella* and its anamorphs, 1. Names published in *Cercospora* and *Passalora*. CBS Biodiversity Series 1. Utrecht, Netherlands.
26. Cruz-Garcia, G.S.; Price, L.L. (2011). Ethnobotanical investigation of “wild” food plants used by rice farmers in Kalasin, Northeast Thailand. *J. Ethnobiol. Ethnomed.* 7, 33.
27. Datchanamurthy, B.; Mythireyi, D.; Divyashanthi, C.M. (2019). Evaluation of antiulcer activity of ethanolic leaf extract of *Coccinia grandis* in indomethacin induced gastric ulcer model. *Int. J. Basic Clin. Pharmacol.* 8(4):629.
28. De Oliveira, J.G.; Vitória, A.P. (2011). Papaya: Nutritional and pharmacological characterization, and quality loss due to physiological disorders. An overview. *Food Res. Int.* 44:1306–1313.
29. Deighton, F.C. (1967). Studies on *Cercospora* and allied genera II. *Passalora*, *Cercosporidium* and some species of *Fusicladium* on *Euphorbia*. *Mycological Papers* 112:1–80.
30. Deighton, F.C. (1971). Studies on *Cercospora* and allied genera III. *Centrospora*. *Mycological Papers.* 124:1–13.
31. Deighton, F.C. (1973). Studies on *Cercospora* and allied genera IV. *Cercosporella* Sacc., *Pseudocercosporella* gen. nov. and *Pseudocercosporidium* gen. nov. *Mycological Papers.* 133:1–62.
32. Deighton, F.C. (1974). Studies on *Cercospora* and allied genera V. *Mycovellosiella* Rangel. and a new species of *Ramulariopsis*. *Mycological Papers.* 137:1–73.
33. Deighton, F.C. (1976). Studies on *Cercospora* and allied genera VI. *Pseudocercospora* Speg., *Pantospora* Cif., and *Cercoseptoria* Petr. *Mycological Papers.* 140:1–168.
34. Deighton, F.C. (1979). Studies on *Cercospora* and allied genera VII. New species and redispositions. *Mycological Papers.* 144:1–56.
35. Deighton, F.C. (1983). Studies on *Cercospora* and allied genera VIII. Further Notes on *Cercoseptoria* and some species and redispositions. *Mycological Papers.* 151:1–13.
36. Deighton, F.C. (1985). Three leaf-spotting hyphomycetes on palms. *Transactions of the British Mycological Society.* 85:739–742.

37. Elizabeth, K. (1994). Immense help from nature's workshop. Ed. 1:207–209.
38. Fatima, S.; Javeed, Z.; Ade, (2006). A. Post-Harvest Rots of Fruits. Discovery Publishing House Delhi, India.
39. Ghani, A. (1957). Medicinal plant of Bangladesh with chemical constituents and uses, 2nd ed. Asiatic Society of Bangladesh, Dhaka. 2003:71-72.
40. Gomes, A.; Das, R.; Sarkhel, S.; Mishra, R.; Mukherjee, S.; Bhattacharya, S.; Gomes, A. (2010). Herbs and herbal constituents active against snake bite. Indian Journal of Experimental Biology. 48:865-878.
41. Hu, T.; Guo, Y.Y.; Zhou, Q.F.; Zhong, X.K.; Zhu, L.; Piao, J.H.; Chen, J.; Jiang, J.G. (2012). Optimization of ultrasonic-assisted extraction of total saponins from *Eclipta prostrata* L. Using response surface methodology. J. Food Sci. 77: C975–C982.
42. Ikan, R.; Ravid, U.; Trosset, D.; Shulman, E. (1971). Ecdysterone: an insect moulting hormone from *Achyranthes aspera* (Amaranthaceae), Cellular and Molecular Life Sciences, 27(5):504-505.
43. Jayalakshmi, V.; Anbumalarmathi, J.; Aruna Sharmili, S. (2019). In vitro Callus induction of *Coccinia indica* (W. and A) and analysis of its phytochemical, antimicrobial, antioxidant and α -amylase inhibitory activity – a comparative study. Int. J. Biochem. Res. Rev. 28 (3):1–16.
44. Jayaraman, P. (1988). Petiole and vein galls of *Achyranthes aspera*, J. Indian bot. Soc. 67:299-301.
45. Jayaweera, D.M.A. (1982). Medicinal used plants in Ceylon, Part IV. Colombo, Sri Lanka, National Science Council of Sri Lanka. 234–236.
46. Kamal (2010). Cercosporoid fungi of India. Bishen Singh Mahendra Pal Singh Publication, Dehradun (Uttarakhand), India. 351 pp. ISBN 10: 8121107539 / ISBN 13: 9788121107532.
47. Kapoor, V.K.; Singh, H. (1966). Isolation of betaine from *Achyranthes aspera* Linn. Indian J. Chem. 4:461-463.
48. Kapoor, V.K.; Singh, H. (1967). Investigation of *Achyranthes aspera* Linn., Indian J. Pharm. 29:285-288.
49. Karunamoorthi, K.; Kim, H.M.; Jegajeevanram, K.; Xavier, J.; Vijayalakshmi, J. (2014). Papaya: A gifted nutraceutical plant—A critical review of recent human health research. Cellmed. 4(2):1–2.17.
50. Khan, A.V.; Khan, A.A. (2006). Ethnomedicinal uses of *Achyranthes aspera* L. (Amaranthaceae) in management of gynecological disorders in western Uttar Pradesh (India). The Journal of Reproductive and Fertility. 43(1):127-129.
51. Kondhare, D.; Lade, H.; (2017). Phytochemical profile, aldose reductase inhibitory, and antioxidant activities of Indian traditional medicinal *Coccinia grandis* (L.) fruit extract. Biotech. 7 (6):378.
52. Krishnaveni, A.; Thaakur, S.R. (2006). Pharmacognostical and preliminary phytochemical studies of *Achyranthes aspera* Linn., Ancient Science of Life.

- 26(1/2):1-5.
53. Kunwar, R.M.; Mahat, L.; Sharma, L.N.; Shrestha, K.P.; Kominee, H.; Bussmann, R.W. (2012). Underutilized plant species in Far West Nepal. *J. Mt. Sci.* 9(5):589–600.
 54. Mahendrarajah, H.; Krishanthiny, K.; Mahendranathan, C. (2020). Antibacterial activity and preliminary screening of phytochemicals of locally available Green Leafy Vegetables in Batticaloa district, Sri Lanka. *AGRIEAST. J. Agric. Sci.* 14(2):9.
 55. Mali, A.; Maheshwari, H.S.; Santani, D.D. (1990). Pharmacological and toxicological activities of alkaloid of *Achyranthes aspera* Linn., *Journal of the Indian Academy of Forensic Sciences.* 29(2):53-58.
 56. Morris, M.J.; Crous, P.W. (1994). New and interesting records of South African fungi. XIV. *Cercosporoid* fungi from weeds. *South African Journal of Botany.* 60:325–332.
 57. Morvin, Yabesh, J.E.; Prabhu, S.; Vijayakumar, S. (2014). An ethnobotanical study of medicinal plants used by traditional healers in silent valley of Kerala, India. *J. Ethnopharmacol.* 154 (3):774–789.
 58. Munir, M.; Qureshi, R. (2018). *Plant and Human Health*, 1. Springer International Publishing Volume 1M. Ozturk & K.R. Hakeem (Eds.) *Plant and Human Health* 1Dm
 59. Nadkarni, A.K. (1954). *Indian Materia Media*, III Ed., I, Popular Book Depot, Bombay, 21.
 60. Neamsuvan, O.; Komonhiran, P.; Boonming, K. (2018). Medicinal plants used for hypertension treatment by folk healers in Songkhla province, Thailand. *J. Ethnopharmacol.* 214:58–70.
 61. Patil, T.; Patil, S.; Patil, A.; Patil, S. (2014). *Carica papaya* leaf extracts—An ethnomedicinal boon. *Int. J. Pharmacogn. Phytochem.* 6:260–265.
 62. Pickering, E.; Steels, E.; Castaneda, R.; Steadman, K.J. (2023). Effect of *Coccinia grandis* (Linn.) Voigt extract on glucose metabolism markers in a prediabetic population: A double-blind randomised clinical trial. *Phytomed, Plus.* 3(4).
 63. Pollack, F.G. (1987). An annotated compilation of *Cercospora* names. *Mycologia Memoir.* 12(1):212.
 64. Pons, N.; Sutton, B.C. (1988). *Cercospora* and similar fungi on yams (*Dioscorea* species). *Mycological Papers.* 160:1–78.
 65. Prasad, S.; Bhattacharya, I.C. (1959). Pharmacognostic study of *Achyranthes aspera* Linn., *Indian Journal of Pharmacy.* 21(3):85.
 66. Rahman, M.; Humaira, T.; & Chowdhury, U. (2023). Bioprospecting of *Coccinia grandis* (L.) Voigt leaf: a wild nutraceutical of Bangladesh. 11(March), 1–8. <https://www.netjournals.org>.
 67. Rahman, M.; Mahmud, T.; Kadir, J.; Abdul Rahman, R.; Begum, M.

- (2008).Major postharvest fungal diseases of papaya cv.'Sekaki'inselangor, malaysia. *Pertanika. J. Trop. Agric. Sci*, 31:27–34.
- 68.Rahman, M.A.; Sarker, J.; Akter, S.; Mamun, A.A.; Azad, M.A.K.; Mohiuddin, M.; Akter, S.; Sarwar, M.S. (2015). Comparative evaluation of antidiabetic activity of crude methanolic extract of leaves, fruits, roots and aerial parts of *Coccinia grandis*. *J. Plant Sci.* 2 (6–1):19–23.
- 69.Raj, S.K.; Snehi, S.K.; Kumar, S.; Pratap, D.; Khan, M.S. (2009). Association of 'Candidatus Phytoplasma asteris' (16SrI group) with yellows of *Achyranthes aspera* in India. *Plant Pathology*. 58:390.
- 70.Raman, A. (2007). Insect-induced plant galls of India: unresolved questions, *Current Science*. 92(6):748-757.
- 71.Reed, C.F.(1976). Information summaries on 1000 economic plants. *Typescripts Submitt. USDA*. 102–103.
- 72.Saeed, F.; Arshad, M.U.; Pasha, I.; Naz, R.; Batool, R.; Khan, A.A.; Nasir, M.A.; Shafique, B. (2014).Nutritional and phyto-therapeutic potential of papaya (*Carica papaya* Linn.): An overview. *Int. J. Food Prop.* 17:1637–1653.
- 73.Selvanayagam, Z.E.; Gnanavendan, S.G.; Balakrishnan, K.; Rao, R.B. (1994).Antisnake venom botanicals from Ethnomedicine. *J. Herbs Spices Med. Plants*. 2:45-100.
- 74.Seshadri, V.; Batta, A.K.; Rangaswami, S. (1981). Structure of two new saponins *Achyranthes aspera*. *Indian J. Chem.* 20B:773-775.
- 75.Sharma, A.; Sharma, R.; Sharma, M.; Kumar, M.; Barbhai, M.D.; Lorenzo, J.M.; Sharma, S.; Samota, M.K.; Atanassova, M.; Caruso, G.(2022).Carica papaya L. Leaves: Deciphering its antioxidant bioactives, biological activities, innovative products, and safety aspects. *Oxid. Med. Cell. Longev.* 1–20.
- 76.Shin, H.D.; Kim, J.D. (2001).Cercospora and allied genera from Korea. *Plant Pathogen from Korea* 7:1–302.
- 77.Shukla, R.; Chakravarty, M.; Gautam, M.P. (2008). Indigenous medicine used for treatment of gynecological disorders by tribal of Chhattisgarh, India. *Journal of Medicinal Plants Research*. 2(12):356-360.
- 78.Singh, V. (1995). Traditional remedies to treat the asthma in the North West and Trans-Himalayan region in J. and K. state. *Fitoterapia*. 66:507-509.
- 79.Sinha, S.K.P.; Dogra, J.V.V. (1985). A survey of plants of Bhagalpur and Santhal pargana for saponin, flavonoids and alkaloids. *Int. J. Crude. Drug. Res.*23:77-86.
- 80.Sinha, Shagun; Navathe, Sudhir; Kharwar, Ravindra;Wijayawardene, Nalin; Dai, Dong-Qin; Chand, Ramesh. (2022). Current status of cercosporoid fungi of India. *Mycotaxon*. 137(2):387.
- 81.Srivastava, N.; Srivastava, A.K.; Kamal. (1994). New synnematos foliicolous hyphomycetes from India. *Mycol. Res.* 98(5):521-524.
- 82.Srivastava, N.; Kamal; Gareth Morgan-Jones. (1995). Notes on Hyphomycetes.

- LXVI. Two new *Ramularia* species from India. MYCOTAXON, LIV: 49-55.
83. Srivastava, N.; Srivastava, A.K.; Kamal. (1995). New hyphopodiate Hyphomycetes from North-Eastern Uttar Pradesh, India. Mycol. Res. 99(4):395-396.
84. Srivastava, N.; Gareth Morgan-Jones. (1996). Notes on Hyphomycetes. LXX. A new species of *Melanographium* from India, with comments on the genus. MYCOTAXON, LVII: 195-200.
85. Srivastava, R.K.; Srivastava, N.; Srivastava A.K. (1994). Additions to genus *Cercospora* from North- Eastern Uttar Pradesh. PROC. NAT. ACAD. SCI., INDIA. 64 (B):I
86. Srivastava, S.; Gupta, M.; Verma, R.K.; Singh S.C.; Kumar, S. (2002). J. Indian Chem. Soc. 79:286.
87. Srivastava, J.; Singh, A. (2022). Diseases of Horticultural Crops: Diagnosis and Management: Volume 1: Fruit Crops; CRC Press: Boca Raton, FL, U.S.A.
88. Srivastava, Mamta; Arya, Mithilesh Kumar; Srivastava, Neeraj. (2011). Cellulolytic fungi causing biodeterioration of Webster's dictionary in Gorakhpur. International Journal of Biological Technology Vol. 2 (Special Issue). 216-220.
89. Srivastava, Neeraj. (2007). Foliicolous hyphomycetous fungi invading plants of Family – Lamiaceae in North-Eastern Uttar Pradesh, India. J. Liv. World. 14(2):78-82.
90. Starley, I.F.; Mohammed, P.; Schneider, G.; Bickler, S.W. (1999). The treatment of paediatric burns using topical papaya. Burns 25:636–639.
91. Thirumalachar, M.J.; Chupp, C. (1948). Notes on some *Cercosporae* from India. Mycologia 40: 352–362.
92. Uddin, M.Z.; Mitu, F.Y.; Rifat, A.B.; Al-Kaium, A. (2019). Ethnomedicinal study focusing on anti-diabetic plants used by the people living in and around Dhaka. Bangladesh J. Plant Taxon. 26(2):231–247.
93. Umer, M.; Mubeen, M.; Iftikhar, Y.; Ali, H.; Zafar-ul-Hye, M.; Asghar, R.; Abbas, M.; Rehman, M.A.; Moya-Elizondo, E.A.; He, Y. (2022). Papaya Ring Spot Virus: An Understanding of a Severe Positive-Sense Single Stranded RNA Viral Disease and its Management. Phyton. 91: 2099–2110.
94. Waisundara, V.Y.; Watawana, M.I.; Jayawardena, N. (2015). *Costus speciosus* and *Coccinia grandis*: traditional medicinal remedies for diabetes. S. Afr. J. Bot. 98, 1–5.
95. Yadav, S.; Pandey, S. and Srivastava, N. (2025). Studies on cellulolytic fungi in Gorakhpur, with special reference to cellulase activity of most frequent fungi. Flora and Fauna. 31(1), 113-120.