

Bioscene Volume- 21 Number- 03 ISSN: 1539-2422 (P) 2055-1583 (O) <u>www.explorebioscene.com</u>

New Report on Antioxidant 'Xanthone' in Some Members of Zingiberaceae

Ishwar Singh¹, Salim¹, Shivani Chauhan^{1*}, Pallavi¹, Karuna¹, Vimala Yerramilli¹, Amit Kumar², Beby Panwar². ^{1,2}Department of Botany, Chaudhary Charan Singh University, Meerut, U.P. India

Corresponding Author: Shivani Chauhan

Abstract

The present investigation is centered on the evaluation of antioxidant activity and detection of antioxidants in different plant parts of some selected members of Zingiberaceae such as Alpinia nutans (L.), Amomum subulatum Roxb., Costus speciosus (Koenig) Sm., Hedychium spicatum Buch. Ham., Elettaria cardamomum Maton., and Curcuma longa (L.). Antioxidant activity was determined by an Assay of POD (peroxidase) activity (EC-1.11.1.7). High phenolic content, higher ascorbic acid content, and high antioxidant activity (POD) in different plant parts of all the selected members were obtained from Alpinia nutans rhizome, Alpinia nutans leaf, and Costus speciosusroot respectively. An attempt has also been made to extract xanthones to confirm the presence of more antioxidant compounds, due to the high antioxidant potential of Zingiberaceae members. Todetect xanthone, methanolic extracts of the plant parts were scanned in the wavelength ranging from 200-800 nm using a Simadzu UV-1800 Series Spectrophotometer, and the characteristic peaks were detected. However, the resulting peaks were observed in the 200-400 nm wavelength. Amomum, Hedychium, and Costus were noted to contain xanthone. This is the first report on the presence of xanthone and the spectrophotometric method for its detection in these plants.

Keywords: antioxidant activity, xanthone, antioxidant, POD, Ascorbic acid,

Zingibers.

Introduction

In Plants, the deleterious consequences of oxidative stress are inhibited or prevented by natural antioxidants which are present in all plants. Plants contain free radical scavengers like polyphenols, flavonoids, and phenolic compounds (Saha & Verma, 2015)Ascorbic acid, also known as vitamin C, is one of the potent naturally occurring antioxidants in a biological system (Evans &Omaye, 2017).The antioxidant property of ascorbic acid is attributed to its ability to reduce potentially damaging reactive oxygen species, forming, resonancestabilized and relatively stable ascorbate free radicals(Buettner, 1993).Plant polyphenols have potent antioxidant properties that's why have the ability to prevent various oxidative stress-associated diseases such as cancer (Taghipour et al., 2017).Therefore, identifying and extracting phenolic compounds from different plants has become a major area of health- and medical-related research. Xanthones arealso a kind of natural product with a polyphenolic structure and of special interest for having many pharmacological effects, such as monoamine oxidase (MAO) inhibition, antitumor, antibacterial, antioxidant, antifungal, anticancer, and anti-inflammatory properties (Negi et al., 2013).It is also reported as an anti-cancer compound (Shan et al., 2011).

The medicinal properties of the rhizomes of Zingiberaceous plants have been widely discussed and accepted worldwide. These plants contain many essential oils which have been reported for their potential antioxidant, anti-inflammatory, and antimicrobial properties (Julie & Ernest, 2012).

Hence, the experimental work presented in this research paper focuses on the screening of some selected members of Zingiberaceae for potential antioxidant activity as well as the detection of antioxidantsin various plant parts. It was expected that through phytochemical evaluation of selected plants, if some new or alternative source of antioxidants is found out, the selected plants may become more useful for humankind.

Materials and Methods

In the present study members screened for the presence of antioxidant compounds were *Alpinia nutans, Costus speciosus, Hedychium spicatum, Zingiber officinale, Elettaria cardamomum, Amomum subulatum Curcuma longa* were selected for the present investigation. The plants have already been nurtured in the Botanical Garden of C.C.S. University Meerut, Uttar Pradesh, India due to their aesthetic, ethical, and medicinal value. Different plant parts such as root, rhizome, shoot, and seed of all the selected members were washed thoroughly with running tap water to remove the entire soil particle. The samples were cut into small pieces, oven-dried at 60°C, ground into fine powder, and stored in airtight polythene bags until use.

Estimation of total phenolic content, ascorbic acid, andPOD activity was carried out by well-defined protocol of (Bray & Thorpe, 1954), (Shukla et al 1979), and (Maehly& Chance 1954) respectively. The standardized protocol of (Liu et al 2010),opted for the extraction of xanthone.

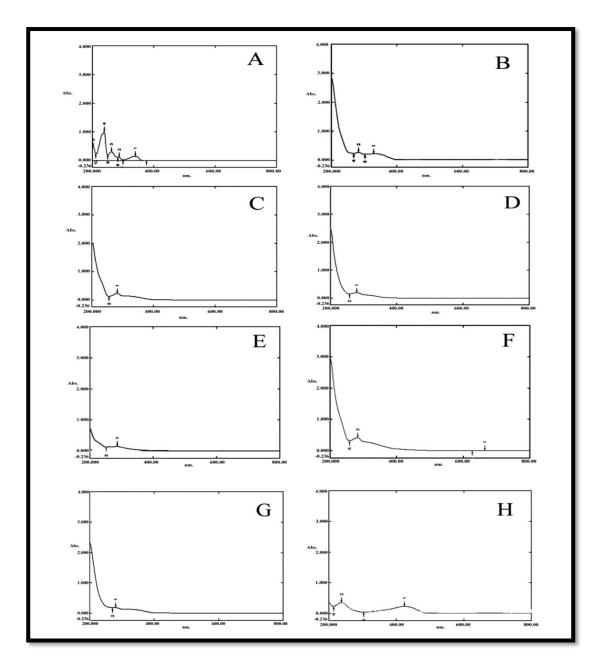
Further analysis for the detection of xanthone was carried outby scanning thesamples' methanolic extract via a spectrophotometer. We find out that the spectrophotometer is a sophisticated instrument for qualitative as well as quantitative analysis of xanthone. A Spectrophotometer for the detection of xanthone is used for the first time in this investigation.

Results

The total phenolic content of selected plant materials ranges from 1.8-24.38mg cinnamic acid eq/g Dwt (dry weight). The ascorbic acid content and POD activity range from 3.15-54.49mg/g Fwt (fresh weight) and 0.27-25.55 A475/min/g Fwt respectively (Table 1 and 2).

Comparison of absorption spectra of standard and sample revealed the presence of xanthone or some polyphenolic compounds in leaf and rhizome of *Alpinia* (Figure 1 B and C), leaf and rhizome of *Amomum* (Figure 1 F and G), rhizome of *Hedychium* (Figure 1 H) and rhizome and seed of *Costus* (Figure 1 D and E). This is not found elsewhere in the literature. No peak was observed in the *Curcuma* leaf, *Curcuma* rhizome, and *Costus* leaf.

Figure 1 (A-H)Spectrophotometric screening for xanthone in plant parts of selected members along with authentic marker (Standard) A- Standard, B-*Alpinia* leaf, C- Alpinia rhizome, D- *Costus* rhizome, E- *Coustus* seed, F-*Amomum* leaf, G- *Amomum* rhizome, H- *Hydechym* rhizome



		PHENOLIC CONTENT, (mg cinnamic acid			
		eq/g Dwt) PLANT PART			
S.No	PLANT NAME	LEAF	RHIZOME	ROOT	SEED
1	Costus speciosus	6.21±0.46	7.29± 0.79	9.39± 0.65	4.82± 3.50
2	Curcuma longa	7.21±0.18	4.7±0.51	23.37± 1.29	NA
3	Hedychiumspicatum	8.93± 0.78	1.8± 0.14	11.95 ± 2.06	NA
4	Alpinia nutans	18.35± 0.54	24.38± 0.92	16.06± 0.69	NA
5	Elettaria cardamomum	6.25± 0.25	15.23± 0.84	13.02± 0.98	NA
6	Amomum subulatum	7.34± 0.61	17.58± 0.96	15.87 ± 0.37	NA

Table- 1 Total phenolic content in different plant parts

NA= Not available

To confirm the presence of more antioxidant compounds due to the high antioxidant potential of members of Zingiberaceae, xanthones were tested using authentic xanthone from Sigma-Aldrich because xanthone is a polyphenolic compound (Sima et al., 2016), which exhibits antioxidant activities as reported by several workers (Cruz et al., 2017; Blanco-Ayala et al., 2013). Spectrophotometric scanning of xanthone was conducted at a wavelength range of 200–400 nm to observe absorption spectra of standard xanthone at different wavelengths.

Plants	Plant Parts	Ascorbic acid (mg/g	POD Activity (\(\Delta A475/min/g)\)	
1 101115		Fwt)	Fwt)	
	Leaf	54.49 ± 1.36	7.49	
Alpinia	Stem	12.99 ± 1.19	1.38	
мрина	Rhizome	10.24 ± 0.10	0.27	
	Root	54.68 ± 0.15	1.11	
	Stem	ND	ND	
a .	Rhizome	7.6 ± 0.16	0.83	
Costus	Root	8.12 ± 0.04	25.55	
	Seed	11.28 ±0.24	6.38	
Hedychiu	Rhizome	4.07 ± 0.54	1.38	
m	Root	9.59 ± 0.89	5.55	
Curcuma Rhizome 3.18		3.15 ± 0.67	1.94	

Table- 2 Ascorbic acid and POD activity in different plant parts

	Root	4.08 ± 0.31	4.16
Zingiber	Rhizome	19.88 ± 0.01	8.88

ND= Not detected

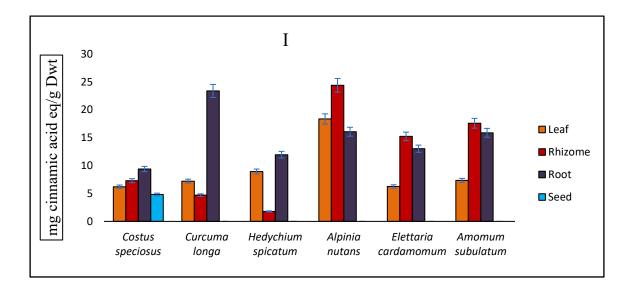
The attribution of the peaks to xanthones was unambiguous, as xanthones present characteristic UV spectra with four bands of decreasing intensity (Liu et al., 2010), However, in our investigation authentic marker exhibited five peaks instead of four with decreasing intensity between 200-400 nm regions of the absorption spectrum (Figure 1A). (Liu et al., 2010), reported that UV spectra of all the xanthone glycosides revealed maximum absorbance near 260 nm. However,we observed that standard xanthone exhibited maximum absorbance at 238.5 nm followed by 202.5 nm (**Table 3**).

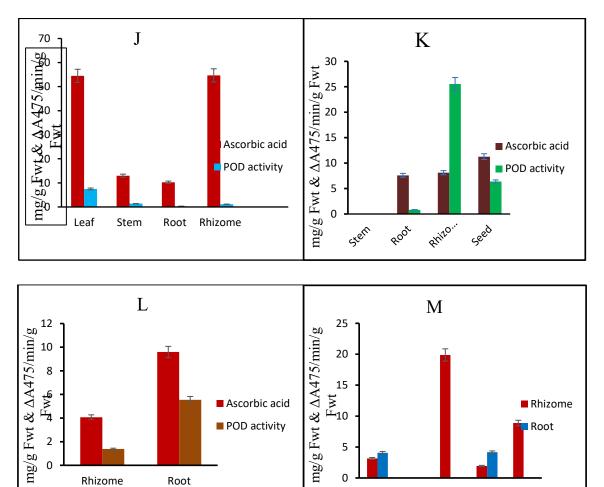
Table- 3 Absorption spectra of standard xanthone at a different wavelength

No.	P/V	Wavelength	Abs.
1	1	339.500	0.153
2	1	287.500	0.123
3	1	262.500	0.304
4	1	238.500	1.017
5	1	202.500	0.460
6	\downarrow	376.000	-0.003
7	\downarrow	300.000	0.024
8	\downarrow	283.000	0.109
9	\downarrow	250.500	0.197
10	\downarrow	211.500	0.233

P=Peak, V=Valley, Upward, and downward arrows show peak and valley respectively

Figure(I-M)graphical representation of Phenolic content, Ascorbic acid, and POD activity in different plant species of Zingiberaceae (I- Phenolic content in different plant parts of members of Zingiberaceae. J- Ascorbic acid and POD activity in different parts of Alpinia, K- Ascorbic acid and POD activity in different parts of Costus, L- Ascorbic acid and POD activity in different parts of Hedychium, M-Ascorbic acid and POD activity in different parts of Curcuma and Zinger.





Discussion

In Alpinia, a leaf with higher antioxidant potential appears to be a better medicinal material than a rhizome, stem, or root. In *Alpinia*, higher phenolic content in stem and rhizome are accompanied by low antioxidant activity (POD) and ascorbic acid content indicating the phenolics to be carrying out the function

Ascorbic acid

POD activity

of defense alone, not the antioxidants, as observed during the study of (Schmidt et al., 2014) where the phenolic compound content increased by more than 100% with solid-state fermentation of rice bran with the *Rhizopus oryzae* fungus and the phenolic extracts of fermented rice bran inhibited DPPH and peroxidase activity. In the other members of Zingiberaceae studied, with only storage parts available, higher phenolic content accompanied by higher ascorbic acid and antioxidant activity (POD) indicating phenolics, ascorbic acid, and antioxidant activity to contribute towards overall antioxidant potential rather than defense of the plant parts. As reported by (Zhang, 2015) phenolic compounds function as antioxidants. Accordingly, we also observed the same relation between POD activity and phenolic compounds and /or antioxidants (Ascorbic acid) as given away in **Tables 1 and 2**.

Conclusion

The potential antioxidant activity as well as the high phenolic content of these selected plantsrevealed that components responsible for the antioxidant activity could be due to the presence of xanthone or some polyphenolic compound.Standardization of the spectrophotometric method for the detection of xanthone will be helpful for quick and cost-effective analysis without using modern techniques.Although further research work for confirmation of xanthone or some other polyphenolic compound using TLC, HPTLC, and FTIR techniques is in progress.

Acknowledgments

Financial assistance as JRF granted to the authors by UGC is acknowledged. The authors thank the Head Department of Botany for providing infrastructural facilities.

References:

- Blanco-Ayala T, Lugo-Huitron R, Serrano-Lopez EM, Reyes-Chilpa R, Rangel-Lopez E, Pineda B, Medina-Campos ON, Sanchez-Chapul L, Pinzon E, Trejo-Solis C, Silva-Adaya D, Pedraza-Chaverri J, Rios C, Perez de la Cruz V, Torres-Ramos M, (2013). Antioxidant properties of xanthones from *Calophyllumbrasiliense:* prevention of oxidative damage induced by FeSO4. BMC Complementary and Alternative Medicince 13: 262.
- 2. Bray HG, Thorpe WV, (1954). Analysis of phenolic compounds of interest in metabolism. Methods Biochemi. Anal. 1: 27-52.
- Buettner GR (1993). The pecking order of free radicals and antioxidants: lipid-peroxidation, alpha-tocopherol, and ascorbate. Arch. Biochem. Biophys. 300: 535–43.

- Cruz I, PuthongkingP, Cravo S, Palmeira A, CidadeH, Pinto M, Sousa E, (2017). Xanthone and Flavone Derivatives as Dual Agents with Acetylcholinesterase Inhibition and Antioxidant Activity as Potential Anti-Alzheimer Agents. Journal of Chemistry. 2017:1-16.
- 5. Evans LW, Omaye ST (2017). Use of Saliva Biomarkers to Monitor Efficacy of Vitamin C in Exercise-Induced Oxidative Stress. Antioxidants. 6(1):5.
- 6. Julie J, Ernest TJ, (2012). Evaluation of the antioxidant potential of rhizome extract of two species of *Alipina*Roxb. (Zingiberaceae). International Research Journal of Pharmacy. 3(4): 402-404.
- Liu X, Liu Y, Chen J, Shi Yan-Ping (2010). Simultaneous Analysis of Xanthone Glycosides in *Halenia elliptica* by HPLC–DAD-ESI-MS. Journal of Chromatographic Science. 48: 76-80.
- 8. Maehly AC, Chance B (1954). The Assay of Catalases and Peroxidases. Methods of Biochemical Analysis. 1: 357-424.
- 9. Negi JS, Bisht VK, Singh P, Rawat MSM, Joshi GP (2013). Naturally Occurring Xanthones: Chemistry and Biology. Journal of Applied Chemistry. 2013: 1-9.
- 10. Saha S, Verma RJ (2015). Antioxidant Activity of Polyphenolic Extract of Phyllanthus emblica against Lead Acetate Induced Oxidative Stress. Toxicology and Environmental Health Sciences. 7(1):82–90.
- 11.Schmidt GC, Goncalves ML, Prietto L, Hackbart HS (2014). Furlong EB. Antioxidant activity and enzyme inhibition of phenolic acids from fermented rice bran with fungus *Rizhopus oryzae*.Food Chemistry. 146, (1) 371-377.
- 12. Shan T, Ma Q, Guo K, Guol K, Liu J, Lil W, Wang F, Wu E, (2011). Xanthones from Mangosteen Extracts as Natural Chemopreventive Agents: Potential Anticancer Drugs. Current molecular medicine. 11(8):666-677.
- 13. Shukla VKS, Kokate CK, Srivastava KC (1979). Spectrometric determination of ascorbic acid. Microchemical Journal. 24: 124-126.
- 14. Sima SYJ, Ng JW, Ng WK, FordeaCG, Henrya CJ, (2016). Plant polyphenols to enhance the nutritional and sensory properties of chocolates. Food Chemistry. 200(1):46-54.
- 15. Taghipour YD, Masoomzadeh S, Farzaei MH, Bahramsoltani R, Karimi-Soureh Z, Rahimi R, Abdollahi M (2017). Polyphenol nanoformulations for cancer therapy: experimental evidence and clinical perspective. International Journal of Nanomedicine. 12: 2689–2702.
- 16. Zhang Yu-Bo, Wu P, Zhang Xiao-Li, Xia C, Li Guo-Qiang, Ye Wen-Cai, Wang Guo-Cai, Li Yao-Lan (2015). Phenolic Compounds from the Flowers of *Bombax malabaricum* and Their Antioxidant and Antiviral Activities. Molecules. 20: 19947–19957.